

Junichi Kurebayashi

Resistance to endocrine therapy in breast cancer

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Abstract Endocrine therapy is the treatment of choice for patients with breast cancer expressing estrogen receptor (ER) and/or progesterone receptor. The efficacy of endocrine therapy is well established in the prevention, adjuvant and metastatic settings. However, either de novo or acquired resistance is frequently observed. Much effort has been made to elucidate the mechanisms of action underlying resistance to endocrine therapy in breast cancer, and several possible explanations have been suggested. Our previous studies have indicated that combined treatment with an antiestrogen, fulvestrant, and an inhibitor of the HER2 signaling pathway, trastuzumab, or an inhibitor of the HER1 signaling pathway, gefitinib, leads to an additive antitumor effect in breast cancer cells expressing ER and HER2 or HER1, respectively. It has also been suggested that the HER1 or HER2 signaling pathway is upregulated during the development of antiestrogen-resistant growth in breast cancer cells. These findings suggest that signal transduction inhibitors are effective for the treatment of antiestrogen-resistant breast cancer. A hypoxic micro-environment has been shown to promote malignant progression in cancer cells. Our previous study and others have suggested that hypoxia posttranscriptionally reduces ER expression and decreases sensitivity to hormonal agents in breast cancer cells. Our preliminary study has also shown that a hypoxic cytotoxin, tirapazamine, increases ER expression in breast cancer xenografts. Differential antitumor activity of tirapazamine on tumor cells under normoxic or hypoxic conditions

may cause this phenomenon. These findings suggest that hypoxic cytotoxins may retard the development of endocrine resistance induced by hypoxia. Molecular mechanisms responsible for endocrine resistance in breast cancer are reviewed and possible therapeutic strategies against this resistance are discussed.

Keywords Resistance · Endocrine therapy · Signal transduction inhibitor · Hypoxia · Hypoxic cytotoxin · Breast cancer

Introduction

Breast cancer, like prostate cancer and endometrial cancer, is classified as one of the hormone-dependent tumors. Estrogen plays important roles in the development and progression of breast cancer. Endocrine therapy, such as estrogen ablation [ovarian ablation or luteinizing hormone-releasing hormone (LH-RH) agonists for premenopausal patients; aromatase inhibitors for postmenopausal patients], antiestrogen therapy or progestin therapy, is the treatment of choice for patients with breast cancer expressing estrogen receptor (ER) and/or progesterone receptor (PgR). The clinical usefulness of endocrine therapy has been proven in the prevention, adjuvant and metastatic settings.

Approximately half of metastatic breast cancer expressing ER and/or PgR responds to endocrine therapy, and postoperative adjuvant endocrine therapy provides approximately a 50% reduction in the development of recurrent disease. These findings suggest that half of breast cancers expressing hormone receptors (HR) are de novo resistant to endocrine therapy. Furthermore, metastatic breast cancer, which initially responds to endocrine therapy, always acquires endocrine resistance (acquired resistance). Both de novo and acquired resistance to endocrine therapy are important problems in the management of breast cancer patients.

A tremendous effort has been made to elucidate the mechanisms of action underlying endocrine resistance in

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J. Kurebayashi
Department of Breast and Thyroid Surgery,
Kawasaki Medical School, 577 Matsushima,
Kurashiki, Okayama 701-0192, Japan
E-mail: kure@med.kawasaki-m.ac.jp
Tel.: +81-86-4621111
Fax: +81-86-4621199

breast cancer over the past two decades. Several possible mechanisms responsible for endocrine resistance have been reported. It should be noted that most of these explanations originated from studies on antiestrogen resistance. The reason for this is because antiestrogens have been most frequently prescribed and antiestrogen resistance has been most frequently observed in clinics. A proportion of antiestrogen-resistant breast cancer subsequently responds to aromatase inhibitors or progestin. However, such breast cancers ultimately acquire resistance to second- and third-line endocrine therapies. These findings suggest that there are multiple steps and complicated mechanisms responsible for endocrine resistance. Novel strategies modifying these steps may be able to retard the development of endocrine resistance and/or overcome endocrine resistance.

Molecular mechanisms responsible for endocrine resistance in breast cancer are reviewed and possible therapeutic strategies against this resistance are discussed.

Mechanisms of action underlying endocrine resistance

Several mechanisms of action have been suggested to be responsible for the development of resistance to endocrine therapy in breast cancer. According to mechanistic points of view, possible mechanisms of action are divided into three categories: (1) reduction in or loss of ER expression; (2) dysfunction of ER signaling; and (3) ligand-independent activation of ER.

Reduction in or loss of ER expression

Endocrine therapy is not suitable for patients with breast cancer expressing no HR, because there is little chance of such breast cancers responding to endocrine therapy. If HR-positive breast cancer loses HR expression during endocrine therapy, such breast cancers become resistant to endocrine therapy. Three possible mechanisms responsible for reduction in ER expression have been reported.

Clonal selection

It is believed that breast cancer consists of estrogen-sensitive and -insensitive clones developing in a mosaic fashion. Endocrine therapy selectively kills HR-positive and estrogen-sensitive clones, while HR-negative and estrogen-insensitive clones grow predominantly. This leads to resistance to endocrine therapy. This mechanism of action appears to be reasonable and a large number of Japanese breast cancer experts believe in this mechanism [60].

Reduction in ER production

Hypermethylation of the promoter region of the ER gene causes transcriptional downregulation and decreases the production of ER protein [40]. Another transcriptional downregulation is caused by a decrease in the expression of transacting factors for ER, such as ER promoter B-associated factor 1 [69]. However, it is not yet known whether these changes directly cause the loss of ER or a loss of ER caused by other mechanisms subsequently induces these changes.

Degradation of ER under hypoxia

An inverse relation between the expression levels of ER and hypoxia-inducible molecules such as hypoxia-inducible factor (HIF)-1 α and carbonic anhydrase IX has been reported in breast cancer tissues [9, 33, 35]. In addition, our previous study and others have suggested that hypoxic conditions lead to posttranscriptional downregulation of ER- α and its function [35]. Subsequently, it was revealed that the reduction in ER expression by a hypoxic environment is mediated by HIF-1 α -driven proteasome-dependent degradation of ER [63]. These findings suggest that hypoxic microenvironments, which are frequently observed in breast tumors, cause a decrease in ER expression and function. It has been suggested that hypoxia promotes malignant progression of many types of human cancers [24]. Therefore, it might be possible that hypoxic microenvironments, which are caused by an increase in tumor size or induced by hormonal agents, provide selective pressure for the outgrowth of ER-poor and more aggressive clones, and finally cause a loss of ER expression in breast tumors.

Although loss of ER in ER-positive tumors is likely to be a main cause for the development of endocrine resistance, several studies have demonstrated that most antiestrogen-resistant breast cancers retain ER expression [30, 46]. These findings indicate that reduction in or loss of ER is not a main cause for the development of endocrine resistance.

Dysfunction of ER signaling

Recent studies have shown that ER-interacting proteins, called co-factors and include co-activators and co-repressors, have important roles in mediating transcriptional activation of target genes by the ER [55]. These findings suggest that quantitative or qualitative changes in ER-related co-factors could contribute to the development of endocrine-resistant breast cancer. However, no definitive relation has been elucidated between alterations of co-activators or co-repressors and endocrine resistance in breast tumors. There are,

however, some preclinical and clinical findings to support these mechanisms.

Quantitative changes in co-factors

Expression of steroid receptor co-activator (SRC)-1 was reported to increase the agonist activity of tamoxifen in experimental cell systems [59]. Additionally, two recent reports have suggested that SRC-1 expression was negatively associated with disease-free survival and positively correlated with human epidermal growth factor receptor (HER) 2 expression in a cohort of breast cancer patients [17, 45]. Similarly, a series of studies has revealed that tamoxifen behaves as an estrogen agonist in breast cancer cells that express high levels of SRC-3 [amplified in breast cancer (AIB)-1] and HER2, and patients whose tumors expressed high levels of both AIB1 and HER2 had worse outcomes with tamoxifen therapy [50, 57].

It was reported that expression of one of the co-repressors, nuclear co-repressor protein (NCOR) 1, was decreased in tamoxifen-resistant tumors in a mouse model system [41]. In addition, low NCOR1 expression was recently reported to be associated with significantly shorter relapse-free survival in breast cancer patients who only received tamoxifen [23].

Qualitative changes in co-factors

A novel co-activator, L7/SPA, has been reported and the complex of this co-activator and tamoxifen was shown to lead to an increase in transcription activation in target genes [65]. Overexpression of this type of co-activator may contribute to endocrine resistance. However, no clinical study has clarified this hypothetical mechanism.

Interaction between ER- α and ER- β or ER- β cx

A recently discovered ER subtype, ER- β , and its isoforms, such as ER- β cx, may play an important role in the development of endocrine resistance. An early series of studies suggested that the mRNA expression levels of wild-type ER- β predict a poorer prognosis and tamoxifen resistance in breast cancer [61, 62]. In contrast, two recent independent studies have proposed that low protein expression levels of ER- β predict resistance to tamoxifen therapy in breast cancer patients [15, 25]. An experimental study suggested that ER- β cx reduces ER- α -mediated transactivation in a dominant-negative fashion [42]. In contrast, a recent study has theorized that protein expression levels of ER- β cx positively correlated with a favorable response to endocrine therapy [51]. The clinical significance of ER- β and its isoforms in the management of breast cancer patients remains controversial and is yet to be determined.

Ligand-independent activation of ER

Classically, transactivation of target genes by ER requires ligand binding to ER. Experimental studies have revealed that several growth factors and their intracellular signaling pathways can stimulate ER activity in the absence of ligand. In addition, it has been suggested that this ligand-independent ER activation is mainly caused by the phosphorylation of different sites in the ER protein [49]. These phenomena may relate to the development of endocrine-resistant breast cancer.

Overexpression of HER1 and/or HER2

It has been reported that HER1 and/or HER2 are overexpressed in 20–30% of primary breast cancers. Clinical studies have suggested that overexpression of HER1 and/or HER2 is related to tamoxifen resistance [26, 47]. Experimental studies have also demonstrated that the intracellular signaling pathway activated by HER1 or HER2, RAS/RAF/MEK/ERK1/2, phosphorylates the serine 118 residue located in the ER protein and leads to ligand-independent transactivation of target genes [6, 31]. Interestingly, it was reported that increased ERK1/2 activity correlates with endocrine resistance and shorter survival in patients with breast cancer [21]. Moreover, experimental studies have suggested that the blockade of these signaling pathways by signal transduction inhibitors may restore the antitumor effect of antiestrogens [54].

Upregulation of AKT activity

It has been proposed that the phosphatidylinositol-3-OH-kinase (PI3K)/AKT (protein kinase B) pathway plays important roles in cell survival and apoptosis [19]. It was reported that phosphorylation of the serine 167 residue located in the ER protein by AKT results in ligand-independent activation of ER [7]. In addition, it has been demonstrated by two different groups that increased AKT1 activation contributes to the aggressive phenotype of tamoxifen-resistant ER-positive breast cancers, and that the overexpression of AKT3 in breast cancer cells results in E2-independent tumor growth; further, the growth of these tumors is enhanced by tamoxifen [16, 64]. It has also been reported that blockage of the AKT signaling pathway by mammalian target of rapamycin (mTOR) inhibition effectively restores the susceptibility of breast cancer cells to tamoxifen [14]. Interestingly, it has been suggested that a negative regulator of AKT, a tumor suppressor PTEN, is frequently inactivated in breast cancer and that there is a strong association between downregulation of PTEN expression in ER-positive tumors and failure of tamoxifen treatment [56].

Upregulation of protein kinase A activity

Protein kinase A (PKA) induced by G-protein-coupled receptors and the 90-kDa ribosomal S6 kinase are also known to be involved in ER phosphorylation [8, 29]. It has recently been reported that phosphorylation of the serine 305 residue in the hinge region of ER- α by PKA induced resistance to tamoxifen, and PKA activity thus induced a switch from antagonistic to agonistic effects of tamoxifen on ER- α [43]. This report also indicated that downregulation of a negative regulator of PKA, PKA-R1 α , was associated with tamoxifen resistance in clinical samples.

Mutations of the ER gene

A missense mutation in the tyrosine 537 residue in the ligand-binding region of ER has been detected in metastatic breast cancer tumors [66]. This mutant ER was reported to be equally active in the absence of ligand or in the presence of estradiol or tamoxifen. Substitution of this residue by different amino acids results in the ligand-independent activation of ER together with co-activator recruitment [67]. However, other studies have suggested that only 1% of primary breast cancers carry gene mutations of ER in the coding region, most of which did not result in alterations at the protein level [53]. Therefore, the clinical significance of ER gene mutations is questionable.

Spliced variants of ER mRNA

An alternatively spliced mRNA lacking specific exons, which encode truncated and potentially constitutively active forms of ER, has been discovered [13]. However, analysis of breast tumors failed to confirm a significant role of the variants in the development of endocrine resistance [4]. The pathological significance of the ER mRNA variants remains to be determined.

Strategies against endocrine-resistant breast cancer

There are many possible mechanisms of action underlying endocrine resistance in breast cancer. Some appear to be irreversible and impossible to overcome. However, others appear to be reversible and might be overcome by adequate strategies. Clinically testable strategies are discussed.

Inhibition of growth factor signaling pathway

It has been suggested that upregulation of growth factor-mediated signaling pathways causes ER phosphorylation, induces ligand-independent ER activation and leads to endocrine-independent and endocrine-resistant

growth of breast cancer cells [49]. Recent clinical studies have also suggested that ER-positive breast cancer overexpressing HER1 and/or HER2 is likely to be resistant to tamoxifen [26, 47]. These findings suggest that inhibitors of growth factor signaling pathways may decrease ER phosphorylation and ligand-independent ER activation, and may re-induce estrogen-dependent growth in breast cancer.

Blockade of HER2 signaling pathway

The humanized anti-HER2 monoclonal antibody, trastuzumab, is the only agent that inhibits the growth factor-mediated signaling pathway and has been introduced into clinical use [58]. Our previous experimental study and others suggested that combined treatment with an antiestrogen and trastuzumab effectively inhibits the growth of breast cancer cells expressing both ER and HER2 [2, 34]. It has been reported that herstatin, a secreted HER2 gene product, which binds to the HER2 ectodomain and blocks HER2 activation, enhanced sensitivity to tamoxifen in breast cancer cells overexpressing HER2 [28]. Inhibition of the mitogen-activated protein kinase (MAPK) pathway driven by HER2 signaling may also be inhibited by MEK inhibitors and this may re-induce tamoxifen sensitivity in breast cancer [39]. Several clinical studies have been launched worldwide to test the clinical usefulness of combined treatment with trastuzumab and endocrine therapeutic agents [68]. However, no synergistic antitumor effect of these combinations has been reported in breast cancer patients.

Blockade of HER1 signaling pathway

A HER1-specific tyrosine kinase inhibitor, gefitinib, was recently introduced as a treatment for nonsmall cell lung cancer [10]. This agent effectively inhibits the signaling pathway mediated by HER1. It has been reported that approximately 20% of breast cancers overexpress HER1, and breast cancer overexpressing HER1 is more aggressive and resistant to endocrine therapy. It has been also suggested that ER-positive breast cancer overexpressing HER1 more rapidly acquires endocrine resistance [3, 38]. Moreover, experimental studies have suggested that gefitinib is not only effective in breast cancer cells overexpressing HER1 but also effective in breast cancer overexpressing HER2 [1]. These findings suggest that gefitinib is a promising agent for combined use with endocrine therapeutic agents. Our previous study and others have suggested that the combined use of gefitinib and an antiestrogen additively inhibited the growth of breast cancer cells expressing both ER and HER1, and that tamoxifen-resistant breast cancer cells associated with HER1 overexpression preferentially responded to this combination therapy [22, 48]. Although limited information is available on the efficacy of gefitinib in the treatment of breast cancer, one preliminary

study suggested that gefitinib was active in patients with tamoxifen-resistant breast cancer [52]. Further clinical studies are needed to clarify the antitumor activity of gefitinib in tamoxifen-resistant breast cancer.

Blockade of other intracellular signaling pathways

Activation of the PI3K/AKT pathway, which is not only driven by HER1/HER2 signaling but also by AKT overexpression or PTEN inactivation, may cause tamoxifen resistance in breast cancer. PKA activity is also suggested to induce a switch from antagonistic to agonistic effects of tamoxifen on ER. These findings suggest that selective blockade of the PI3K/AKT or PKA pathways may overcome tamoxifen resistance in breast cancer [14, 43].

Retardation of the development of acquired endocrine resistance

Even if it is difficult to overcome endocrine resistance, it may be possible to prolong the antitumor effect of endocrine therapy or retard the development of endocrine resistance. Two promising strategies are discussed.

Intermittent, alternating and combined endocrine therapies

Acquired endocrine resistance has been recognized as an adaptive response of breast cancer cells to endocrine therapy. Consequently, intermittent treatment with a single endocrine therapeutic agent, alternating use of two different agents, or combined treatment with two different agents may interfere with the adaptation process and retard the development of endocrine resistance.

Prostate-specific antigen is a sensitive and specific tumor marker. A new strategy in which intermittent treatment with a LH-RH agonist is given to patients with advanced prostate cancer has been tested in a clinical trial [70]. Unfortunately, there is no such sensitive and specific tumor marker in the management of breast cancer. However, approximately 70% of metastatic breast cancer patients have at least one elevated tumor marker, such as CA 15-3, CEA or NCC-ST-439 [37]. If a certain endocrine therapy is effective and an elevated tumor marker decreases during the treatment of metastatic breast cancer patients, endocrine therapy can be stopped until the tumor marker increases again. This intermittent treatment may prolong the efficacy of endocrine therapy.

Recent clinical studies have demonstrated that alternating treatment with tamoxifen and an aromatase inhibitor is more effective than tamoxifen alone in the adjuvant setting for postmenopausal patients with early breast cancer [5, 11]. It is possible that the sequential

administration of an aromatase inhibitor effectively destroys breast cancer cells acquiring tamoxifen resistance. However, the mechanisms of action underlying the efficacy of alternating treatment of two different endocrine therapies remain to be elucidated.

Concurrent use of tamoxifen and a LH-RH agonist has been reported to be more effective than respective agents alone in premenopausal patients with metastatic breast cancer [32]. Recent clinical studies have also indicated that this concurrent use is as effective as chemotherapy, such as the cyclophosphamide, methotrexate, fluorouracil (CMF) regimen, in the adjuvant setting for premenopausal breast cancer patients [27]. It has recently been reported that concurrent use of a third-generation aromatase inhibitor and a LH-RH agonist provides a high rate of clinical benefit to premenopausal metastatic breast cancer patients [18]. Concurrent use of an aromatase inhibitor and a LH-RH agonist is also under investigation in the adjuvant setting for premenopausal breast cancer patients [44].

Targeting hypoxic tumor cells

Our previous study and others have suggested that an epigenetic change, such as a hypoxic microenvironment, may play an important role in the development of endocrine resistance in breast cancer [12, 35, 63]. If so, hypoxic cytotoxins, such as tirapazamine which selectively kills cancer cells under hypoxic conditions [20], may be useful for retarding the development of endocrine resistance. Our preliminary study suggested that the administration of hypoxic cytotoxins restored ER expression in human breast cancer xenografts transplanted into athymic nude mice [36]. It can be speculated that the hypoxic cytotoxin destroyed hypoxic breast cancer cells with a low level of ER, and that normoxic breast cancer cells with a high level of ER preferentially survived in the xenografts. If this is the case, normoxic tumor cells should be responsive to endocrine therapy. Further experimental studies are clearly needed to clarify this interesting phenomenon.

Conclusions

Although there are many possible mechanisms of action responsible for the development of endocrine resistance, a common cascade has not yet been discovered. In other words, there appears to be no common cascade. Multi-step or complicated mechanisms may be involved in the development of endocrine resistance. Generally, endocrine therapy is less toxic and provides better quality of life to patients with breast cancer in comparison with cytotoxic chemotherapy. Therefore, clinical researchers have to make a continuous effort to develop new strategies for enhancing or prolonging the efficacy of endocrine therapy.

In consideration with clinical implications, strategies for retarding endocrine resistance, such as intermittent, alternating or combined endocrine therapy, are testable in clinical trials. In addition, concomitant or sequential administration of signal transduction inhibitors, such as trastuzumab or gefitinib, may not only prolong the efficacy of endocrine therapy but also overcome a part of endocrine resistance in the near future. Well-designed clinical trials supported by scientific rationale are essential to clarify the usefulness of novel strategies as described above.

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References

- Anido J, Matar P, Albanell J, Guzman M, Rojo F, Arribas J, Averbuch S, Baselga J (2003) ZD1839, a specific epidermal growth factor receptor (EGFR) tyrosine kinase inhibitor, induces the formation of inactive EGFR/HER2 and EGFR/HER3 heterodimers and prevents heregulin signaling in HER2-overexpressing breast cancer cells. *Clin Cancer Res* 9:1274–1283
- Argiris A, Wang CX, Whalen SG, DiGiovanna MP (2004) Synergistic interactions between tamoxifen and trastuzumab (Herceptin). *Clin Cancer Res* 10:1409–1420
- Arpino G, Green SJ, Allred DC, Lew D, Martino S, Osborne CK, Ellledge RM (2004) HER-2 amplification, HER-1 expression, and tamoxifen response in estrogen receptor-positive metastatic breast cancer: a southwest oncology group study. *Clin Cancer Res* 10:5670–5676
- Balleine RL, Hunt SM, Clarke CL (1999) Coexpression of alternatively spliced estrogen and progesterone receptor transcripts in human breast cancer. *J Clin Endocrinol Metab* 84:1370–1377
- Boccardo F, Rubagotti A, Amoroso D, Mesiti M, Romeo D, Caroti C, Farris A, Cruciani G, Villa E, Schieppati G, Mustacchi G; Italian Breast Cancer Cooperative Group (2001) Sequential tamoxifen and aminoglutethimide versus tamoxifen alone in the adjuvant treatment of postmenopausal breast cancer patients: results of an Italian cooperative study. *J Clin Oncol* 19:4209–4215
- Bunone G, Briand PA, Miksicek RJ, Picard D (1996) Activation of the unliganded estrogen receptor by EGF involves the MAP kinase pathway and direct phosphorylation. *EMBO J* 15:2174–2183
- Campbell RA, Bhat-Nakshatri P, Patel NM, Constantinidou D, Ali S, Nakshatri H (2001) Phosphatidylinositol 3-kinase/AKT-mediated activation of estrogen receptor alpha: a new model for anti-estrogen resistance. *J Biol Chem* 276:9817–9824
- Chen D, Pace PE, Coombes RC, Ali S (1999) Phosphorylation of human estrogen receptor alpha by protein kinase A regulates dimerization. *Mol Cell Biol* 19:1002–1015
- Chen SK, Wykoff CC, Watson PH, Han C, Leek RD, Pastorek J, Gatter KC, Ratcliffe P, Harris AL (2001) Prognostic significance of a novel hypoxia-regulated marker, carbonic anhydrase IX, in invasive breast carcinoma. *J Clin Oncol* 19:3660–3668
- Cohen MH, Williams GA, Sridhara R, Chen G, McGuinn WD Jr, Morse D, Abraham S, Rahman A, Liang C, Lostritto R, Baird A, Pazdur R (2004) United States food and drug administration drug approval summary: Gefitinib (ZD1839; Iressa) tablets. *Clin Cancer Res* 10:1212–1218
- Coombes RC, Hall E, Gibson LJ, Paridaens R, Jassem J, Delozier T, Jones SE, Alvarez I, Bertelli G, Ortmann O, Coates AS, Bajetta E, Dodwell D, Coleman RE, Fallowfield LJ, Mickiewicz E, Andersen J, Lonning PE, Cocconi G, Stewart A, Stuart N, Snowdon CF, Carpentieri M, Massimini G, Bliss JM; Intergroup Exemestane Study (2004) A randomized trial of exemestane after two to three years of tamoxifen therapy in postmenopausal women with primary breast cancer. *N Engl J Med* 350:1081–1092
- Cooper C, Liu GY, Niu YL, Santos S, Murphy LC, Watson PH (2004) Intermittent hypoxia induces proteasome-dependent down-regulation of estrogen receptor alpha in human breast carcinoma. *Clin Cancer Res* 10:8720–8727
- Daffada AA, Johnston SR, Smith IE, Detre S, King N, Dowsett M (1995) Exon 5 deletion variant estrogen receptor messenger RNA expression in relation to tamoxifen resistance and progesterone receptor/pS2 status in human breast cancer. *Cancer Res* 55:288–293
- deGraffenried LA, Friedrichs WE, Russell DH, Donzis EJ, Middleton AK, Silva JM, Roth RA, Hidalgo M (2004) Inhibition of mTOR activity restores tamoxifen response in breast cancer cells with aberrant Akt activity. *Clin Cancer Res* 10:8059–8067
- Esslimani-Sahla M, Simony-Lafontaine J, Kramar A, Lavail R, Mollevi C, Warner M, Gustafsson JA, Rochefort H (2004) Estrogen receptor beta (ER beta) level but not its ER beta cx variant helps to predict tamoxifen resistance in breast cancer. *Clin Cancer Res* 10:5769–5776
- Faridi J, Wang L, Endemann G, Roth RA (2003) Expression of constitutively active Akt-3 in MCF-7 breast cancer cells reverses the estrogen and tamoxifen responsiveness of these cells in vivo. *Clin Cancer Res* 9:2933–2939
- Fleming FJ, Myers E, Kelly G, Crotty TB, McDermott EW, O'Higgins NJ, Hill AD, Young LS (2004) Expression of SRC-1, AIB1, and PEA3 in HER2 mediated endocrine resistant breast cancer; a predictive role for SRC-1. *J Clin Pathol* 57:1069–1074
- Forward DP, Cheung KL, Jackson L, Robertson JF (2004) Clinical and endocrine data for goserelin plus anastrozole as second-line endocrine therapy for premenopausal advanced breast cancer. *Br J Cancer* 90:590–594
- Franke TF, Hornik CP, Segev L, Shostak GA, Sugimoto C (2003) PI3K/Akt and apoptosis: size matters. *Oncogene* 22:8983–8998
- Gandara DR, Lara PN Jr, Goldberg Z, Le QT, Mack PC, Lau DH, Gumerlock PH (2002) Tirapazamine: prototype for a novel class of therapeutic agents targeting tumor hypoxia. *Semin Oncol* 29 (1 Suppl 4):102–109
- Gee JM, Robertson JF, Ellis IO, Nicholson RI (2001) Phosphorylation of ERK1/2 mitogen-activated protein kinase is associated with poor response to anti-hormonal therapy and decreased patient survival in clinical breast cancer. *Int J Cancer* 95:247–254
- Gee JM, Harper ME, Hutcheson IR, Madden TA, Barrow D, Knowlden JM, McClelland RA, Jordan N, Wakeling AE, Nicholson RI (2003) The anti-epidermal growth factor receptor agent gefitinib (ZD1839/Iressa) improves anti-hormone response and prevents development of resistance in breast cancer in vitro. *Endocrinology* 144:5105–5117
- Girault I, Lerebours F, Amarir S, Tozlu S, Tubiana-Hulin M, Lidereau R, Bieche I (2003) Expression analysis of estrogen receptor alpha coregulators in breast carcinoma: evidence that NCOR1 expression is predictive of the response to tamoxifen. *Clin Cancer Res* 9:1259–1266
- Hockel M, Vaupel P (2001) Tumor hypoxia: definitions and current clinical, biologic, and molecular aspects. *J Natl Cancer Inst* 93:266–276
- Hopp TA, Weiss HL, Parra IS, Cui Y, Osborne CK, Fuqua SA (2004) Low levels of estrogen receptor beta protein predict resistance to tamoxifen therapy in breast cancer. *Clin Cancer Res* 10:7490–7499

26. Houston SJ, Plunkett TA, Barnes DM, Smith P, Rubens RD, Miles DW (1999) Overexpression of c-erbB2 is an independent marker of resistance to endocrine therapy in advanced breast cancer. *Br J Cancer* 79:1220–1226
27. Jakesz R, Hausmaninger H, Kubista E, Gnant M, Menzel C, Bauernhofer T, Seifert M, Haider K, Mlineritsch B, Steindorfer P, Kwasny W, Fridrik M, Steger G, Wette V, Samonigg H; Austrian Breast and Colorectal Cancer Study Group Trial 5 (2002) Randomized adjuvant trial of tamoxifen and goserelin versus cyclophosphamide, methotrexate, and fluorouracil: evidence for the superiority of treatment with endocrine blockade in premenopausal patients with hormone-responsive breast cancer—Austrian Breast and Colorectal Cancer Study Group Trial 5. *J Clin Oncol* 20:4621–4627
28. Jhabvala-Romero F, Evans A, Guo S, Denton M, Clinton GM (2003) Herstatin inhibits heregulin-mediated breast cancer cell growth and overcomes tamoxifen resistance in breast cancer cells that overexpress HER-2. *Oncogene* 22:8178–8186
29. Joel PB, Smith J, Sturgill TW, Fisher TL, Blenis J, Lannigan DA (1998) pp90rsk1 regulates estrogen receptor-mediated transcription through phosphorylation of Ser-167. *Mol Cell Biol* 18:1978–1984
30. Johnston SR, Saccani-Jotti G, Smith IE, Salter J, Newby J, Coppen M, Ebbs SR, Dowsett M (1995) Changes in estrogen receptor, progesterone receptor, and pS2 expression in tamoxifen-resistant human breast cancer. *Cancer Res* 55:3331–3338
31. Kato S, Endoh H, Masuhiro Y, Kitamoto T, Uchiyama S, Sasaki H, Masushige S, Gotoh Y, Nishida E, Kawashima H (1995) Activation of the estrogen receptor through phosphorylation by mitogen-activated protein kinase. *Science* 270:1491–1494
32. Klijn JG, Beex LV, Mauriac L, van Zijl JA, Veyret C, Wildiers J, Jassem J, Piccart M, Burghouts J, Becquart D, Seynaeve C, Mignolet F, Duchateau L (2000) Combined treatment with buserelin and tamoxifen in premenopausal metastatic breast cancer: a randomized study. *J Natl Cancer Inst* 92:903–911
33. Kronblad A, Helczynska K, Nielsen NH, Pahlman E, Emdin S, Pahlman S, Landberg G (2003) Regional cyclin D1 overexpression or hypoxia correlate inversely with heterogeneous oestrogen receptor- α expression in human breast cancer. *In Vivo* 17:311–318
34. Kunisue H, Kurebayashi J, Otsuki T, Tang CK, Kurosumi M, Yamamoto S, Tanaka K, Doihara H, Shimizu N, Sonoo H (2000) Anti-HER2 antibody enhances the growth inhibitory effect of anti-oestrogen on breast cancer cells expressing both oestrogen receptors and HER2. *Br J Cancer* 82:46–51
35. Kurebayashi J, Otsuki T, Moriya T, Sonoo H (2001) Hypoxia reduces hormone responsiveness of human breast cancer cells. *Jpn J Cancer Res* 92:1093–1101
36. Kurebayashi J, Yamamoto Y, Okubo S, Sonoo H, Nagasawa H, Uto Y, Hori H, Moriya T (2003) Experimental study for overcoming endocrine resistance in breast cancer: implication of hypoxic cytotoxins (in Japanese). *Basic Invest Breast Carcinoma* 12:35–38
37. Kurebayashi J, Nishimura R, Tanaka K, Kohno N, Kurosumi M, Moriya T, Ogawa Y, Taguchi T (2004) Significance of serum tumor markers in monitoring advanced breast cancer patients treated with systemic therapy: a prospective study. *Breast Cancer* 11:389–395
38. Kurebayashi J, Okubo S, Yamamoto Y, Sonoo H (2004) Inhibition of HER1 signaling pathway enhances antitumor effect of endocrine therapy in breast cancer. *Breast Cancer* 11:38–41
39. Kurokawa H, Lenferink AE, Simpson JF, Pisacane PI, Sliwkowski MX, Forbes JT, Arteaga CL (2000) Inhibition of HER2/neu (erbB-2) and mitogen-activated protein kinases enhances tamoxifen action against HER2-overexpressing, tamoxifen-resistant breast cancer cells. *Cancer Res* 60:5887–5894
40. Lapidus RG, Nass SJ, Butash KA, Parl FF, Weitzman SA, Graff JG, Herman JG, Davidson NE (1998) Mapping of ER gene CpG island methylation-specific polymerase chain reaction. *Cancer Res* 58:2515–2519
41. Lavinsky RM, Jepsen K, Heinzel T, Torchia J, Mullen TM, Schiff R, Del-Rio AL, Ricote M, Ngo S, Gensch J, Hilsenbeck SG, Osborne CK, Glass CK, Rosenfeld MG, Rose DW (1998) Diverse signaling pathways modulate nuclear receptor recruitment of N-CoR and SMRT complexes. *Proc Natl Acad Sci USA* 95:2920–2925
42. Maruyama K, Endoh H, Sasaki-Iwaoka H, Kanou H, Shimaya E, Hashimoto S, Kato S, Kawashima H (1998) A novel isoform of rat estrogen receptor beta with 18 amino acid insertion in the ligand binding domain as a putative dominant negative regulator of estrogen action. *Biochem Biophys Res Commun* 246:142–147
43. Michalides R, Griekspoor A, Balkenende A, Verwoerd D, Janssen L, Jalink K, Floore A, Velds A, van't Veer L, Neeffjes J (2004) Tamoxifen resistance by a conformational arrest of the estrogen receptor alpha after PKA activation in breast cancer. *Cancer Cell* 5:597–605
44. Michaud LB, Jones KL, Buzdar AU (2001) Combination endocrine therapy in the management of breast cancer. *Oncologist* 6:538–546
45. Myers E, Fleming FJ, Crotty TB, Kelly G, McDermott EW, O'higgins NJ, Hill AD, Young LS (2004) Inverse relationship between ER- β and SRC-1 predicts outcome in endocrine-resistant breast cancer. *Br J Cancer* 91:1687–1693
46. Newby JC, Johnston SR, Smith IE, Dowsett M (1997) Expression of epidermal growth factor receptor and c-erbB2 during the development of tamoxifen resistance in human breast cancer. *Clin Cancer Res* 3:1643–1651
47. Nicholson RI, McClelland RA, Gee JM, Manning DL, Cannon P, Robertson JF, Ellis IO, Blamey RW (1994) Epidermal growth factor receptor expression in breast cancer: association with response to endocrine therapy. *Breast Cancer Res Treat* 29:117–125
48. Okubo S, Kurebayashi J, Otsuki T, Yamamoto Y, Tanaka K, Sonoo H (2004) Additive antitumour effect of the epidermal growth factor receptor tyrosine kinase inhibitor gefitinib (Iressa, ZD1839) and the antioestrogen fulvestrant (Faslodex, ICI 182,780) in breast cancer cells. *Br J Cancer* 90:236–244
49. Osborne CK, Schiff R, Fuqua SA, Shou J (2001) Estrogen receptor: current understanding of its activation and modulation. *Clin Cancer Res* 7(12 Suppl):4338s–4342s
50. Osborne CK, Bardou V, Hopp TA, Chamness GC, Hilsenbeck SG, Fuqua SA, Wong J, Allred DC, Clark GM, Schiff R (2003) Role of the estrogen receptor coactivator AIB1 (SRC-3) and HER-2/neu in tamoxifen resistance in breast cancer. *J Natl Cancer Inst* 95:353–361
51. Palmieri C, Lam EW, Mansi J, MacDonald C, Shousha S, Madden P, Omoto Y, Sunters A, Warner M, Gustafsson JA, Coombes RC (2004) The expression of ER beta cx in human breast cancer and the relationship to endocrine therapy and survival. *Clin Cancer Res* 10:2421–2428
52. Robertson JFR, Gutteridge E, Cheung KL, Oweres R, Koehler M, Hamilton L (2002) A phase II study of ZD1839 (Iressa) in tamoxifen resistant ER positive and endocrine insensitive (ER negative) breast cancer. *Breast Cancer Res Treat* 76:S96
53. Roodi N, Bailey LR, Kao WY, Verrier CS, Yee CJ, Dupont WD, Parl FF (1995) Estrogen receptor gene analysis in estrogen receptor-positive and receptor-negative primary breast cancer. *J Natl Cancer Inst* 87:446–451
54. Schiff R, Massarweh SA, Shou J, Bharwani L, Mohsin SK, Osborne CK (2004) Cross-talk between estrogen receptor and growth factor pathways as a molecular target for overcoming endocrine resistance. *Clin Cancer Res* 10 (1 Pt 2):331S–336S
55. Shibata H, Spencer TE, Onate SA, Jenster G, Tsai SY, Tsai MJ, O'Malley BW (1997) Role of co-activators and co-repressors in the mechanism of steroid/thyroid receptor action. *Recent Prog Horm Res* 52:141–164

56. Shoman N, Klassen S, McFadden A, Bickis MG, Torlakovic E, Chibbar R (2005) Reduced PTEN expression predicts relapse in patients with breast carcinoma treated by tamoxifen. *Mod Pathol* 18:250–259
57. Shou J, Massarweh S, Osborne CK, Wakeling AE, Ali S, Weiss H, Schiff R (2004) Mechanisms of tamoxifen resistance: increased estrogen receptor-HER2/neu cross-talk in ER/HER2-positive breast cancer. *J Natl Cancer Inst* 96:926–935
58. Slamon DJ, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A, Fleming T, Eiermann W, Wolter J, Pegram M, Baselga J, Norton L (2001) Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *N Engl J Med* 344:783–792
59. Smith CL, Nawaz Z, O'Malley BW (1997) Coactivator and corepressor regulation of the agonist/antagonist activity of the mixed antiestrogen, 4-hydroxytamoxifen. *Mol Endocrinol* 11:657–666
60. Sonoo H, Kurebayashi J, Iino Y, Inaji H, Watanabe T, Toi M, Kobayashi S, Sato B, Yoshimoto M (1999) Current status and controversial issues concerning endocrine therapy for patients with recurrent breast cancer in Japan. *Breast Cancer* 6:344–350
61. Speirs V, Malone C, Walton DS, Kerin MJ, Atkin SL (1999) Increased expression of estrogen receptor beta mRNA in tamoxifen-resistant breast cancer patients. *Cancer Res* 59:5421–5424
62. Speirs V, Parkes AT, Kerin MJ, Walton DS, Carleton PJ, Fox JN, Atkin SL (1999) Coexpression of estrogen receptor alpha and beta: poor prognostic factors in human breast cancer? *Cancer Res* 59:525–528
63. Stoner M, Saville B, Wormke M, Dean D, Burghardt R, Safe S (2002) Hypoxia induces proteasome-dependent degradation of estrogen receptor alpha in ZR-75 breast cancer cells. *Mol Endocrinol* 16:2231–2242
64. Sun M, Paciga JE, Feldman RI, Yuan Z, Coppola D, Lu YY, Shelley SA, Nicosia SV, Cheng JQ (2001) Phosphatidylinositol-3-OH Kinase (PI3K)/AKT2, activated in breast cancer, regulates and is induced by estrogen receptor alpha (ERalpha) via interaction between ERalpha and PI3K. *Cancer Res* 61:5985–5991
65. Takimoto GS, Graham JD, Jackson TA, Tung L, Powell RL, Horwitz LD, Horwitz KB (1999) Tamoxifen resistant breast cancer: coregulators determine the direction of transcription by antagonist-occupied steroid receptors. *J Steroid Biochem Mol Biol* 69:45–50
66. Weis KE, Ekena K, Thomas JA, Lazennec G, Katzenellenbogen BS (1996) Constitutively active human estrogen receptors containing amino acid substitutions for tyrosine 537 in the receptor protein. *Mol Endocrinol* 10:1388–1398
67. White R, Sjöberg M, Kalkhoven E, Parker MG (1997) Ligand-independent activation of the oestrogen receptor by mutation of a conserved tyrosine. *EMBO J* 16:1427–1435
68. Winer EP, Burstein HJ (2001) New combinations with Herceptin in metastatic breast cancer. *Oncology* 61:50–57
69. Yoshida T, Eguchi H, Nakachi K, Tanimoto K, Higashi Y, Suemasu K, Iino Y, Morishita Y, Hayashi S (2000) Distinct mechanisms of loss of estrogen receptor alpha gene expression in human breast cancer: methylation of the gene and alteration of trans-acting factors. *Carcinogenesis* 21:2193–2201
70. Youssef E, Tekyi-Mensah S, Hart K, Bolton S, Forman J (2003) Intermittent androgen deprivation for patients with recurrent/metastatic prostate cancer. *Am J Clin Oncol* 26:e119–e123